



A guideline to limit indoor airborne transmission of COVID-19

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Edited by Renyi Zhang, Texas A&M University, College Station, TX, and accepted by Editorial Board Member John H. Seinfeld March 3, 2021 (received for review September 9, 2020)

The current revival of the American economy is being predicated on social distancing, specifically the Six-Foot Rule, a guideline that offers little protection from pathogen-bearing aerosol droplets sufficiently small to be continuously mixed through an indoor space. The importance of airborne transmission of COVID-19 is now widely recognized. While tools for risk assessment have recently been developed, no safety guideline has been proposed to protect against it. We here build on models of airborne disease transmission in order to derive an indoor safety guideline that would impose an upper bound on the “cumulative exposure time,” the product of the number of occupants and their time in an enclosed space. We demonstrate how this bound depends on the rates of ventilation and air filtration, dimensions of the room, breathing rate, respiratory activity and face mask use of its occupants, and infectiousness of the respiratory aerosols. By synthesizing available data from the best-characterized indoor spreading events with respiratory drop size distributions, we estimate an infectious dose on the order of 10 aerosol-borne virions. The new virus (severe acute respiratory syndrome coronavirus 2 [SARS-CoV-2]) is thus inferred to be an order of magnitude more infectious than its forerunner (SARS-CoV), consistent with the pandemic status achieved by COVID-19. Case studies are presented for classrooms and nursing homes, and a spreadsheet and online app are provided to facilitate use of our guideline. Implications for contact tracing and quarantining are considered, and appropriate caveats enumerated. Particular consideration is given to respiratory jets, which may substantially elevate risk when face masks are not worn.

COVID-19 | infectious aerosol | airborne transmission | SARS-CoV-2 coronavirus | indoor safety guideline

COVID-19 is an infectious pneumonia that appeared in Wuhan, Hubei Province, China, in December 2019 and has since caused a global pandemic (1, 2). The pathogen responsible for COVID-19, severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), is known to be transported by respiratory droplets exhaled by an infected person (3–7). There are thought to be three possible routes of human-to-human transmission of COVID-19: large drop transmission from the mouth of an infected person to the mouth, nose or eyes of the recipient; physical contact with droplets deposited on surfaces (fomites) and subsequent transfer to the recipient’s respiratory mucosae; and inhalation of the microdroplets ejected by an infected person and held aloft by ambient air currents (6, 8). We subsequently refer to these three modes of transmission as, respectively, “large-drop,” “contact,” and “airborne” transmission, while noting that the distinction between large-drop and airborne transmission is somewhat nebulous given the continuum of sizes of emitted droplets (11).^{*} We here build upon the existing theoretical framework for describing airborne disease transmission (12–18) in order to characterize the evolution of the concentration of pathogen-laden droplets in a well-mixed room, and the associated risk of infection to its occupants.

The Six-Foot Rule is a social distancing recommendation by the US Centers for Disease Control and Prevention, based on

the assumption that the primary vector of pathogen transmission is the large drops ejected from the most vigorous exhalation events, coughing and sneezing (5, 19). Indeed, high-speed visualization of such events reveals that 6 ft corresponds roughly to the maximum range of the largest, millimeter-scale drops (20). Compliance to the Six-Foot Rule will thus substantially reduce the risk of such large-drop transmission. However, the liquid drops expelled by respiratory events are known to span a considerable range of scales, with radii varying from fractions of a micron to millimeters (11, 21).

There is now overwhelming evidence that indoor airborne transmission associated with relatively small, micron-scale aerosol droplets plays a dominant role in the spread of COVID-19 (4, 5, 7, 17–19, 22), especially for so-called “superspreading events” (25–28), which invariably occur indoors (29). For example, at the 2.5-h-long Skagit Valley Chorale choir practice that took place in Washington State on March 10, some 53 of 61 attendees were infected, presumably not all of them within 6 ft of the initially infected individual (25). Similarly, when 23 of 68 passengers were infected on a 2-h bus journey in Ningbo, China, their seated locations were uncorrelated with distance to the index case (28). Airborne transmission was also implicated in the COVID-19 outbreak between residents of a Korean high-rise building whose apartments were linked via air ducts (30). Studies have also confirmed the presence of infectious SARS-CoV-2

Significance

Airborne transmission arises through the inhalation of aerosol droplets exhaled by an infected person and is now thought to be the primary transmission route of COVID-19. By assuming that the respiratory droplets are mixed uniformly through an indoor space, we derive a simple safety guideline for mitigating airborne transmission that would impose an upper bound on the product of the number of occupants and their time spent in a room. Our theoretical model quantifies the extent to which transmission risk is reduced in large rooms with high air exchange rates, increased for more vigorous respiratory activities, and dramatically reduced by the use of face masks. Consideration of a number of outbreaks yields self-consistent estimates for the infectiousness of the new coronavirus.

Author contributions: M.Z.B. and J.W.M.B. designed research; M.Z.B. and J.W.M.B. performed research; M.Z.B. analyzed data; and M.Z.B. and J.W.M.B. wrote the paper.

The authors declare no competing interest.

This article is a PNAS Direct Submission. R.Z. is a guest editor invited by the Editorial Board.

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This article contains supporting information online at <https://www.pnas.org/lookup/suppl/doi:10.1073/pnas.2018995118/-DCSupplemental>.

Published April 13, 2021.

^{*}The possibility of pathogen resuspension from contaminated surfaces has also recently been explored (9, 10).

virions in respiratory aerosols (31) suspended in air samples collected at distances as large as 16 ft from infected patients in a hospital room (3). Further evidence for the dominance of indoor airborne transmission has come from an analysis of 7,324 early cases outside the Hubei Province, in 320 cities across mainland China (32). The authors found that all clusters of three or more cases occurred indoors, 80% arising inside apartment homes and 34% potentially involving public transportation; only a single transmission was recorded outdoors. Finally, the fact that face mask directives have been more effective than either lockdowns or social distancing in controlling the spread of COVID-19 (22, 33) is consistent with indoor airborne transmission as the primary driver of the global pandemic.

The theoretical model developed herein informs the risk of airborne transmission resulting from the inhalation of small, aerosol droplets that remain suspended for extended periods within closed, well-mixed indoor spaces. When people cough, sneeze, sing, speak, or breathe, they expel an array of liquid droplets formed by the shear-induced or capillary destabilization of the mucosal linings of the lungs and respiratory tract (8, 34, 35) and saliva in the mouth (36, 37). When the person is infectious, these droplets of sputum are potentially pathogen bearing, and represent the principle vector of disease transmission. The range of the exhaled pathogens is determined by the radii of the carrier droplets, which typically lie in the range of 0.1 μm to 1 mm. While the majority are submicron in scale, the drop size distribution depends on the form of exhalation event (11). For normal breathing, the drop radii vary between 0.1 and 5.0 μm , with a peak around 0.5 μm (11, 38, 39). Relatively large drops are more prevalent in the case of more violent expiratory events such as coughing and sneezing (20, 40). The ultimate fate of the droplets is determined by their size and the airflows they encounter (41, 42). Exhalation events are accompanied by a time-dependent gas-phase flow emitted from the mouth that may be roughly characterized in terms of either continuous turbulent jets or discrete puffs (20, 38, 43). The precise form of the gas flow depends on the nature of the exhalation event, specifically the time dependence of the flux of air expelled. Coughs and sneezes result in violent, episodic puff releases (20), while speaking and singing result in a puff train that may be well approximated as a continuous turbulent jet (38, 43). Eventually, the small droplets settle out of such turbulent gas flows. In the presence of a quiescent ambient, they then settle to the floor; however, in the well-mixed ambient more typical of a ventilated space, sufficiently small drops may be suspended by the ambient airflow and mixed throughout the room until being removed by the ventilation outflow or inhaled (*SI Appendix, section 1*).

Theoretical models of airborne disease transmission in closed, well-mixed spaces are based on the seminal work of Wells (44) and Riley et al. (45), and have been applied to describe the spread of airborne pathogens including tuberculosis, measles, influenza, H1N1, coronavirus (SARS-CoV) (12–16, 46, 47), and, most recently, the novel coronavirus (SARS-CoV-2) (17, 25). These models are all based on the premise that the space of interest is well mixed; thus, the pathogen is distributed uniformly throughout. In such well-mixed spaces, one is no safer from airborne pathogens at 60 ft than 6 ft. The Wells–Riley model (13, 15) highlights the role of the room’s ventilation outflow rate Q in the rate of infection, showing that the transmission rate is inversely proportional to Q , a trend supported by data on the spreading of airborne respiratory diseases on college campuses (48). The additional effects of viral deactivation, sedimentation dynamics, and the polydispersity of the suspended droplets were considered by Nicas et al. (14) and Stilianakis and Drossinos (16). The equations describing pathogen transport in well-mixed, closed spaces are thus well established and have recently been applied to provide risk assessments for indoor

airborne COVID-19 transmission (17, 18). We use a similar mathematical framework here in order to derive a simple safety guideline.

We begin by describing the dynamics of airborne pathogen in a well-mixed room, on the basis of which we deduce an estimate for the rate of inhalation of pathogen by its occupants. We proceed by deducing the associated infection rate from a single infected individual to a susceptible person. We illustrate how the model’s epidemiological parameter, a measure of the infectiousness of COVID-19, may be estimated from available epidemiological data, including transmission rates in a number of spreading events, and expiratory drop size distributions (11). Our estimates for this parameter are consistent with the pandemic status of COVID-19 in that they exceed those of SARS-CoV (17); however, our study calls for refined estimates through consideration of more such field data. Most importantly, our study yields a safety guideline for mitigating airborne transmission via limitation of indoor occupancy and exposure time, a guideline that allows for a simple quantitative assessment of risk in various settings. Finally, we consider the additional risk associated with respiratory jets, which may be considerable when face masks are not being worn.

The Well-Mixed Room

We first characterize the evolution of the pathogen concentration in a well-mixed room. The assumption of well mixedness is widely applied in the theoretical modeling of indoor airborne transmission (14, 16, 17), and its range of validity is discussed in *SI Appendix, section 1*. We describe the evolution of the airborne pathogen by adapting standard methods developed in chemical engineering to describe the “continuously stirred tank reactor” (49), as detailed in *SI Appendix, section 1*. We assume that the droplet-borne pathogen remains airborne for some time before being extracted by the room’s ventilation system, inhaled, or sedimenting out. The fate of ejected droplets in a well-mixed ambient is determined by the relative magnitudes of two speeds: the settling speed of the drop in quiescent air, v_s , and the ambient air circulation speed within the room, v_a . Drops of radius $r \leq 100 \mu\text{m}$ and density ρ_d descend through quiescent air of density ρ_a and dynamic viscosity μ_a at the Stokes settling speed $v_s(r) = 2\Delta\rho gr^2/(9\mu_a)$, prescribed by the balance between gravity and viscous drag (50), where g is the gravitational acceleration and $\Delta\rho = \rho_d - \rho_a$.

We consider a well-mixed room of area A , depth H , and volume $V = HA$ with ventilation outflow rate Q and outdoor air change rate (typically reported as air changes per hour, or ACH) $\lambda_a = Q/V$. Mechanical ventilation imposes an additional recirculation flow rate Q_r that further contributes to the well-mixed state of the room, but alters the emergent drop size distributions only if accompanied by filtration. The mean air velocity, $v_a = (Q + Q_r)/A$, prescribes the air mixing time, $\tau_a = H/v_a = H^2/(2D_a)$, where $D_a = v_a H/2$ is the turbulent diffusivity defined in terms of the largest eddies (51, 52), those on the scale of the room (53). The timescale of the droplet settling from a well-mixed ambient corresponds to that through a quiescent ambient (51, 52, 54), as justified in *SI Appendix, section 1*. Equating the characteristic times of droplet settling, H/v_s , and removal, V/Q , indicates a critical drop radius $r_c = \sqrt{9\lambda_a H \mu_a / (2g\Delta\rho)}$ above which drops generally sediment out, and below which they remain largely suspended within the room prior to removal by ventilation outflow. We here define airborne transmission as that associated with droplets with radius $r < r_c$. The relevant physical picture, of particles settling from a well-mixed environment, is commonly invoked in the contexts of stirred aerosols (51) and sedimentation in geophysics (54). The additional effects of ventilation, particle dispersity, and pathogen deactivation in the context of airborne disease transmission were considered by Nicas et al. (14), Stilianakis and Drossinos (16)

and Buonanno et al. (17, 18), whose models will be built upon here.

In *SI Appendix, section 1*, we provide justification for our assumption of the well-mixed room. It is noteworthy that, even in the absence of forced ventilation, there will generally be some mixing in an enclosed space: Natural ventilation will lead to flows through windows and doors, as well as leakage through construction materials and joints. Moreover, occupants serve to enhance airflow through their motion and respiration. Traditionally, ventilation standards for American homes (American Society of Heating, Refrigerating and Air-Conditioning Engineers [ASHRAE]) recommend a minimal outdoor air exchange rate of $\lambda_a = 0.35/\text{h}$, a value comparable to the average of $0.34/\text{h}$ reported for Chinese apartments, including those in winter in Wuhan (55). Even with such minimal ventilation rates, for a room of height $H = 2.1$ m, there is an associated critical drop size of radius $r_c = 1.3$ μm . In order to guard against infectious aerosols, ASHRAE now recommends ventilation rates greater than $\lambda_a = 6/\text{h}$, which corresponds to $r_c = 5.5$ μm . The “airborne” droplets of interest here, those of radius $r < r_c$, thus constitute a significant fraction of those emitted in most respiratory events (11, 23, 38).

Wells (56) argued that exhaled drops with diameter less than approximately 100 μm will evaporate before settling. The resulting “droplet nuclei” consist of residual solutes, including dissolved salts, carbohydrates, proteins, and pathogens, which are typically hygroscopic and retain significant quantities of bound water (57, 58). For a droplet with initial radius r_0 , the equilibrium size, $r_{eq} = r_0 \sqrt[3]{\phi_s/(1-RH)}$, is reached over an evaporation timescale, $\tau_e = r_0^2/(\theta(1-RH))$, where ϕ_s is the initial solute volume fraction, RH is the relative humidity, and $\theta = 4.2 \times 10^{-10}$ m^2/s at 25°C (58). In dry air ($RH \ll 1$), saliva droplets, which typically contain 0.5% solutes and a similar volume of bound water ($\phi_s \approx 1\%$), can thus lose up to $1 - \sqrt[3]{0.01} \approx 80\%$ of their initial size (58). Conversely, droplets of airway mucus shrink by as little as $1 - \sqrt[3]{0.2} \approx 40\%$, since they typically contain 5 to 10% gel-forming mucins (glycosylated proteins) and comparable amounts of bound water (59). The evaporation time at 50% RH ranges from $\tau_e = 1.2$ ms for $r_0 = 0.5$ μm to 12 s at 50 μm . These inferences are consistent with experiments demonstrating that stable respiratory aerosol distributions in the range $r_{eq} < 10$ μm are reached within 0.8 s of exhalation (11). While we note that the drop size distributions will, in general, depend on the relative humidity, we proceed by employing the equilibrium drop distributions measured directly (11, 38).

We consider a polydisperse suspension of exhaled droplets characterized by the number density $n_d(r)$ (per volume of air, per radius) of drops of radius r and volume $V_d(r) = 4/3\pi r^3$. The drop size distribution $n_d(r)$ is known to vary strongly with respiratory activity and various physiological factors (11, 17, 39). The drops contain a microscopic pathogen concentration $c_v(r)$, a drop size-dependent probability of finding individual virions (3, 31, 60), usually taken to be that in the sputum (RNA copies per milliliter) (17, 61).

The virions become deactivated (noninfectious) at a rate $\lambda_v(r)$ that generally depends on droplet radius, temperature, and humidity (62). Using data for human influenza viruses (63), a roughly linear relationship between λ_v and RH can be inferred (62, 64), which provides some rationale for the seasonal variation of flu outbreaks, specifically, the decrease from humid summers to dry winters. Recent experiments on the aerosol viability of model viruses (bacteriophages) by Lin and Marr (65) have further revealed a nonmonotonic dependence of λ_v on relative humidity. Specifically, the deactivation rate peaks at intermediate values of relative humidity, where the cumulative exposure of virions to disinfecting salts and solutes is maximized. Since

the dependence $\lambda_v(RH)$ is not yet well characterized experimentally for SARS-CoV-2, we follow Miller et al. (25) and treat the deactivation rate as bounded by existing data, specifically, $\lambda_v = 0$ [no deactivation measured in 16 h at $22 \pm 1^\circ\text{C}$ and $RH = 53 \pm 11\%$ (66)] and $\lambda_v = 0.63/\text{h}$ [corresponding to a half life of 1.1 h at $23 \pm 2^\circ\text{C}$ and $RH = 65\%$ (67)]. Pending further data for SARS-CoV-2, we assume $\lambda_v = 0.6RH$ h^{-1} , and note the rough consistency of this estimate with that for MERS-CoV (Middle East Respiratory Syndrome coronavirus) at 25°C and $RH = 79\%$ (68), $\lambda_v = 1.0/\text{h}$. Finally, we note that effective viral deactivation rates may be enhanced using either ultraviolet radiation (UV-C) (69) or chemical disinfectants (e.g. H_2O_2 , O_3) (70).

The influence of air filtration and droplet settling in ventilation ducts may be incorporated by augmenting $\lambda_v(r)$ by an amount $\lambda_f(r) = p_f(r)\lambda_r$, where $p_f(r)$ is the probability of droplet filtration and $\lambda_r = Q_r/V$. The recirculation flow rate, Q_r , is commonly expressed in terms of the primary outdoor air fraction, $Z_p = Q/(Q_r + Q)$, where $Q + Q_r$ is the total airflow rate. We note that the United States Environmental Protection Agency defines high-efficiency particulate air (HEPA) filtration (71) as that characterized by $p_f > 99.97\%$ for aerosol particles. Ordinary air filters have required Minimum Efficiency Reporting Value (MERV) ratings of $p_f = 20$ to 90% in specific size ranges. Other types of filtration devices (22), such as electrostatic precipitators (72) with characteristic p_f values of 45 to 70%, can also be included in this framework.

We seek to characterize the concentration $C(r, t)$ (specifically, number/volume per radius) of pathogen transported by drops of radius r . We assume that each of $I(t)$ infectious individuals exhales pathogen-laden droplets of radius r at a constant rate $P(r) = Q_b n_d(r) V_d(r) p_m(r) c_v(r)$ (number/time per radius), where Q_b is the breathing flow rate (exhaled volume per time). We introduce a mask penetration factor, $0 < p_m(r) < 1$, that roughly accounts for the ability of masks to filter droplets as a function of drop size (73–76).[†] The concentration, $C(r, t)$, of pathogen suspended within drops of radius r then evolves according to

$$V \frac{\partial C}{\partial t} = I P - (Q + p_f Q_r + v_s A + \lambda_v V) C \quad [1]$$

Rate of Change = Production rate from exhalation - Loss rate from ventilation, filtration, sedimentation, and deactivation

where $v_s(r)$ is the particle settling speed and $p_f(r)$ is, again, the probability of drop filtration in the recirculation flow Q_r . Owing to the dependence of the settling speed on particle radius, the population of each drop size evolves, according to Eq. 1, at different rates. Two limiting cases of Eq. 1 are of interest. For the case of $\lambda_v = v_s = Q_r = 0$, drops of infinitesimal size that are neither deactivated nor removed by filtration, it reduces to the Wells–Riley model (44, 45). For the case of $\lambda_v = P = Q = Q_r = 0$, a nonreacting suspension with no ventilation, it corresponds to established models of sedimentation from a well-mixed ambient (51, 54). For the sake of notational simplicity, we define a size-dependent sedimentation rate $\lambda_s(r) = v_s(r)/H = \lambda_a(r/r_c)^2$ as the inverse of the time taken for a drop of radius r to sediment from ceiling to floor in a quiescent room.

When one infected individual enters a room at time $t = 0$, so that $I(0) = 1$, the radius-resolved pathogen concentration increases as $C(r, t) = C_s(r) (1 - e^{-\lambda_c(r)t})$, relaxing to a steady value, $C_s(r) = P(r)/(\lambda_c(r)V)$, at a rate $\lambda_c(r) = \lambda_a + \lambda_f(r) +$

[†] For the sake of simplicity, we do not consider here the dependence of p_m on respiratory activity (77) or direction of airflow (78), but note that, once reliably characterized, these dependencies might be included in a straightforward fashion.

$\lambda_s(r) + \lambda_v(r)$. Note that both the equilibrium concentration and the timescale to approach it are decreased by the combined effects of ventilation, air filtration, particle settling, and deactivation (14, 64). Owing to the dependence of this adjustment process on the drop size, one may understand it as a dynamic sifting process wherein larger droplets settle out and reach their equilibrium concentration relatively quickly. However, we note that, in the absence of filtration and deactivation ($\lambda_f = \lambda_v = 0$), the adjustment time, λ_c^{-1} , depends only weakly on drop size, varying from $V/(2Q)$ for the largest airborne drops (with radius r_c) to V/Q for infinitesimal drops. The sedimentation rate of the “airborne” droplets of radius $r \leq r_c$ is thus bounded above by the air exchange rate, $\lambda_s(r) \leq \lambda_a$. The exhaled drop size distribution depends strongly on respiratory activity (11, 17, 38, 39); thus, so too must the radius-resolved concentration of airborne pathogen. The predicted dependence on respiratory activity (11) of the steady-state volume fraction of airborne droplets, $\phi_s(r) = C_s(r)/c_v(r)$, is illustrated in Fig. 1.

We define the airborne disease transmission rate, $\beta_a(t)$, as the mean number of transmissions per time per infectious individual per susceptible individual. One expects $\beta_a(t)$ to be proportional to the quantity of pathogen exhaled by the infected person, and to that inhaled by the susceptible person. Gammaitoni and Nucci (12) defined the airborne transmission rate as $\beta_a(t) = Q_b c_i C_s(t)$ for the case of a population evolving according to the Wells–Riley model and inhaling a monodisperse suspension. Here, c_i is the viral infectivity, the parameter that connects the fluid physics to the epidemiology, specifically, the concentration of suspended pathogen to the infection rate. We note its relation to the notion of “infection quanta” in the epidemiological literature (44). Specifically, $c_i < 1$ is the infection quanta per pathogen, while $c_i^{-1} > 1$ is the “infectious dose,” the number of aerosol-borne virions required to cause infection with probability $1 - e^{-1} = 63\%$.

For the polydisperse suspension of interest here, we define the airborne transmission rate as

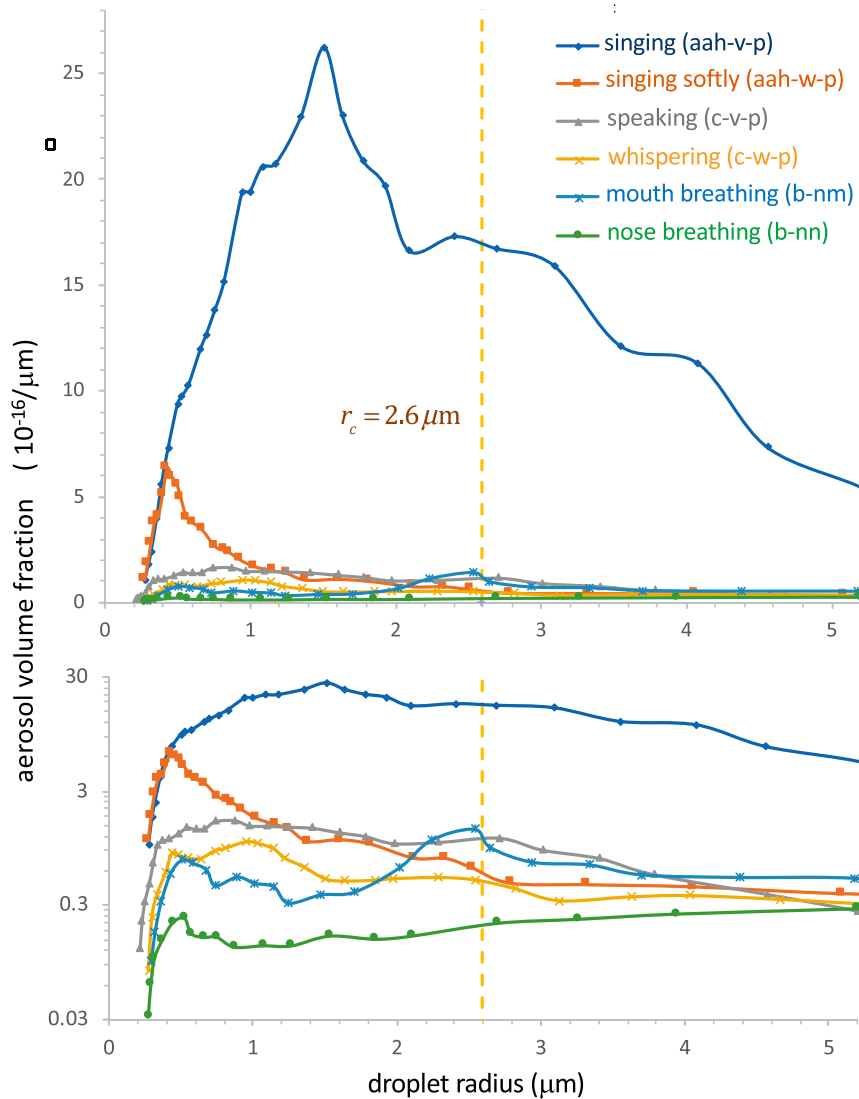


Fig. 1. Model predictions for the steady-state, droplet radius-resolved aerosol volume fraction, $\phi_s(r)$, produced by a single infectious person in a well-mixed room. The model accounts for the effects of ventilation, pathogen deactivation, and droplet settling for several different types of respiration in the absence of face masks ($p_m = 1$). The ambient conditions are taken to be those of the Skagit Valley Chorale superspreading incident (25, 27) ($H = 4.5$ m, $A = 180$ m², $\lambda_a = 0.65$ h⁻¹, $r_c = 2.6$ μ m, $\lambda_v = 0.3$ h⁻¹, and $RH = 50\%$). The expiratory droplet size distributions are computed from the data of Morawska et al. (ref. 11, figure 3) at $RH = 59.4\%$ for aerosol concentration per log-diameter, using $n_d(r) = (dC/d \log D)/(r \ln 10)$. The breathing flow rate is assumed to be 0.5 m³/h for nose and mouth breathing, 0.75 m³/h for whispering and speaking, and 1.0 m³/h for singing.

$$\beta_a(t) = Q_b s_r \int_0^\infty C(r, t) p_m(r) c_i(r) dr, \quad [2]$$

thereby accounting for the protective properties of masks, and allowing for the possibility that the infectivity $c_i(r)$ depends on droplet size. Different droplet sizes may emerge from, and penetrate into, different regions of the respiratory tract (34, 37, 79), and so have different $c_i(r)$; moreover, virions in relatively small droplets may diffuse to surfaces more rapidly and so exchange with bodily fluids more effectively. Such a size dependence in infectivity, $c_i(r)$, is also consistent with reports of enhanced viral shedding in micron-scale aerosols compared to larger drops for both influenza virus (60) and SARS-CoV-2 (31). Finally, we introduce a relative transmissibility (or susceptibility), s_r , to rescale the transmission rate for different subpopulations or viral strains.

Indoor Safety Guideline

The reproduction number of an epidemic, \mathcal{R}_0 , is defined as the mean number of transmissions per infected individual. Provided $\mathcal{R}_0 < 1$, a disease will not spread at the population level (80). Estimates of \mathcal{R}_0 for COVID-19 have been used to compare its rate of spread in different regions and its dependence on different control strategies (33, 81–83) and, most recently, viral variants (84, 85). We here define an analogous reproductive number for indoor, airborne transmission, $\mathcal{R}_{in}(\tau)$, as the expected number of transmissions in a room of total occupancy N over a time τ from a single infected person entering at $t = 0$.

Our safety guideline sets a small risk tolerance ϵ (typically 1 to 10%) for the indoor reproductive number, defined as

$$\mathcal{R}_{in}(\tau) = N_s \int_0^\tau \beta_a(t) dt < \epsilon. \quad [3]$$

The number of susceptibles, $N_s = p_s(N - 1)$, may include all others in the room ($p_s = 1$), or be reduced by the susceptible probability $p_s < 1$, the fraction of the local population not yet exposed or immunized. In the limit of $\epsilon \ll 1$, one may interpret $\mathcal{R}_{in}(\tau)$ as the probability of the first transmission, which is approximately equal to the sum of the N_s independent probabilities of transmission to any particular susceptible individual in a well-mixed room.[‡] In *SI Appendix, section 3*, we show that this guideline follows from standard epidemiological models, including the Wells–Riley model, but note that it has broader generality. The exact transient safety bound appropriate for the time-dependent situation arising directly after an infected index case enters a room is evaluated in *SI Appendix, section 2*.

We here focus on a simpler, more conservative guideline that follows for long times relative to the air residence time, $\tau \gg \lambda_a^{-1}$ (which may vary from minutes to hours, and is necessarily greater than $\lambda_c(r)^{-1}$), when the airborne pathogen has attained its equilibrium concentration $C(r, t) \rightarrow C_s(r)$. In this equilibrium case, the transmission rate (2) becomes constant,

$$\bar{\beta}_a = \frac{Q_b^2 p_m^2}{V} \int_0^\infty \frac{n_q(r)}{\lambda_c(r)} dr = \frac{Q_b^2 p_m^2}{V} \frac{C_q}{\lambda_c(\bar{r})} = p_m^2 f_d \lambda_q, \quad [4]$$

where, for the sake of simplicity, we assume constant mask filtration p_m over the entire range of aerosol drop sizes. We define the microscopic concentration of infection quanta per liquid volume as $n_q(r) = n_d(r) V_d(r) c_v(r) c_i(r)$, and the concentration of infection quanta or “infectiousness” of exhaled air,

$C_q = \int_0^\infty n_q(r) dr$. The latter is the key disease-specific parameter in our model, which can also be expressed as the rate of quanta emission by an infected person, $\lambda_q = Q_b C_q$. The second equality in Eq. 4 defines the effective infectious drop radius \bar{r} , given in *SI Appendix, Eq. S7*. The third equality defines the dilution factor, $f_d = Q_b / (\lambda_c(\bar{r}) V)$, the ratio of the concentration of infection quanta in the well-mixed room to that in the unfiltered breath of an infected person. As we shall see in what follows, f_d provides a valuable diagnostic in assessing the relative risk of various forms of exposure.

We thus arrive at a simple guideline, appropriate for steady-state situations, that bounds the cumulative exposure time (CET),

$$(N - 1)\tau < \epsilon \frac{\bar{\lambda}_c V + \bar{v}_s A}{Q_b^2 p_m^2 C_q s_r}. \quad [5]$$

where $\bar{v}_s = v_s(\bar{r})$, and $\bar{\lambda}_c = \lambda_a + \lambda_f(\bar{r}) + \lambda_v(\bar{r})$ is the air purification rate associated with air exchange, air filtration, and viral deactivation. The effect of relative humidity on the droplet size distribution can be captured by multiplying \bar{r} by $\sqrt[3]{0.4/(1 - RH)}$, since the droplet distributions used in our analysis were measured at $RH = 60\%$ (11).

By noting that the sedimentation rate of aerosols is usually less than the air exchange rate, $\lambda_s(r) < \lambda_a$, and by neglecting the influence of both air filtration and pathogen deactivation, we deduce, from Eq. 5, a more conservative bound on the CET,

$$N\tau < \epsilon \frac{\lambda_a V}{Q_b^2 p_m^2 C_q s_r}, \quad [6]$$

the interpretation of which is immediately clear. To minimize risk of infection, one should avoid spending extended periods in highly populated areas. One is safer in rooms with large volume and high ventilation rates. One is at greater risk in rooms where people are exerting themselves in such a way as to increase their respiration rate and pathogen output, for example, by exercising, singing, or shouting. Since the rate of inhalation of contagion depends on the volume flux of both the exhalation of the infected individual and the inhalation of the susceptible person, the risk of infection increases as Q_b^2 . Likewise, masks worn by both infected and susceptible persons will reduce the risk of transmission by a factor p_m^2 , a dramatic effect given that $p_m \leq 0.1$ for moderately high-quality masks (74, 75).

Application to COVID-19

The only poorly constrained quantity in our guideline is the epidemiological parameter, $C_q s_r$, the product of the concentration of exhaled infection quanta by an infectious individual, C_q , and the relative transmissibility, s_r . We emphasize that C_q and s_r are expected to vary widely between different populations (86–91), among individuals during progression of the disease (92, 93), and between different viral strains (84, 85). Nevertheless, we proceed by making rough estimates for C_q for different respiratory activities on the basis of existing epidemiological data gathered from early superspreading events of COVID-19. Our inferences provide a baseline value for C_q , relevant for elderly individuals exposed to the original strain of SARS-CoV-2, that we may rescale by the relative transmissibility s_r in order to consider different populations and viral strains. We make these inferences with the hope that such an attempt will motivate the acquisition of more such data, and so lead to improved estimates for C_q and s_r for different populations in various settings.

An inference of $C_q = 970$ quanta/m³ was made by Miller et al. (25) in their recent analysis of the Skagit Valley Chorale superspreading incident (27), on the basis of the assumption that the transmission was described in terms of the Wells–Riley model (12, 13, 17, 45). To be precise, they inferred a quanta

[‡]Markov's inequality ensures that the probability of at least one transmission, P_1 , is bounded above by the expected number of transmissions, $P_1 \leq R_{in}$. In the limit, $R_{in} < \epsilon \ll 1$, these quantities are asymptotically equal, since $P_1 = 1 - (1 - p(\tau))^{N_s} \sim N_s p(\tau) = R_{in}$ for N_s independent transmissions of probability, $p(\tau) = \int_0^\tau \beta_a(t) dt \ll 1$.

emission rate of $\lambda_q = C_q \bar{Q}_b = 970$ quanta/h for a mean breathing rate of $\bar{Q}_b = 1.0$ m³/h appropriate for singing (25). This inference is roughly consistent with studies of other related viral diseases. For example, Liao et al. (46) estimated $C_q = 28$ quanta/m³ from the rate of indoor spreading of SARS-CoV, in a hospital and an elementary school. Estimates of C_q for H1N1 influenza fall in the range 15 to 128 quanta/m³ (47). For SARS-CoV-2, Buonanno et al. (17) estimate a C_q range of 10.5 to 1,030 quanta/m³, on the basis of the estimated infectivity $c_i = 0.01$ to 0.1 of SARS-CoV (94) and the reported viral loads in sputum (92, 93, 95), and note that the precise value depends strongly on the infected person's respiratory activity. Notably, their range spans the high value inferred for the Skagit Valley Chorale (25), and all of our inferences to follow.

We proceed by estimating quanta concentrations, C_q , or, equivalently, quanta emission rates, $\lambda_q = \bar{Q}_b C_q$, for different forms of respiration. First, we solve Eq. 1 to obtain the steady-state radius-resolved droplet volume fraction $\phi_s(r)$ for various hypothetical expiratory activities in the room of the Skagit Valley Chorale, using the drop size distributions of Morawska et al. (11). Our results are shown in Fig. 1. Integrating each curve up to the critical radius r_c , we then obtain an activity-dependent volume fraction of infectious airborne droplets $\phi_1 = \int_0^{r_c} \phi_s(r) dr$ in the choir room (see *SI Appendix*). Finally, we assume the inferred value, $C_q = 970$ quanta/m³, for the superspreading incident (25) that resulted from the expiratory activity most resembling singing [voiced "aahs" with pauses for recovery (11)], and deduce values of C_q for other forms of respiration by rescaling with the appropriate ϕ_1 values. Our predictions for the dependence of C_q on respiratory activity are shown in Fig. 2. For validation, we also show estimates for C_q based on the recent measurements

of activity-dependent aerosol concentrations reported by Asadi et al. (38, 39). Specifically, we calculated the aerosol volume fractions from the reported drop-size distributions (from figure 5 of ref. 39) for a different set of expiratory activities that included various breathing patterns and speaking aloud at different volumes. We then used these volume fractions to rescale the value $C_q = 72$ quanta/m³ for speaking at intermediate volume (39), which we chose to match the value inferred for the most similar respiratory activity considered by Morawska et al. (11), specifically, voiced counting with pauses (11). Notably, the quanta concentrations so inferred, C_q , are consistent across the full range of activities, from nasal breathing at rest (1 to 10 quanta/m³) to oral breathing and whispering (5 to 40 quanta/m³), to loud speaking and singing (100 to 1,000 quanta/m³).

Our inferences for C_q from a number of superspreading events are also roughly consistent with physiological measurements of viral RNA in the bodily fluids of COVID-19 patients at peak viral load. Specifically, our estimate of $C_q = 72$ quanta/m³ for voiced counting (11) and intermediate-volume speech (39) with integrated aerosol volume fractions $\phi_1 = 0.36$ and 0.11 ($\mu\text{m}/\text{cm}^3$) corresponds, respectively, to microscopic concentrations of $c_q = c_i c_v = 2 \times 10^8$ and 7×10^8 quanta/mL (see *SI Appendix*). Respiratory aerosols mainly consist of sputum produced by the fragmentation (96) of mucous plugs and films in the bronchioles and larynx (34–36). Larger droplets are thought to form by fragmentation of saliva in the mouth (36, 37). Airborne viral loads are usually estimated from that of saliva or sputum (61, 92, 93, 95, 97). After incubation, viral loads, c_v , in sputum tend to peak in the range 10^8 to 10^{11} RNA copies per milliliter (92, 93, 95), while much lower values have been reported for other bodily fluids (92, 93, 98). Virus shedding in the pharynx remains

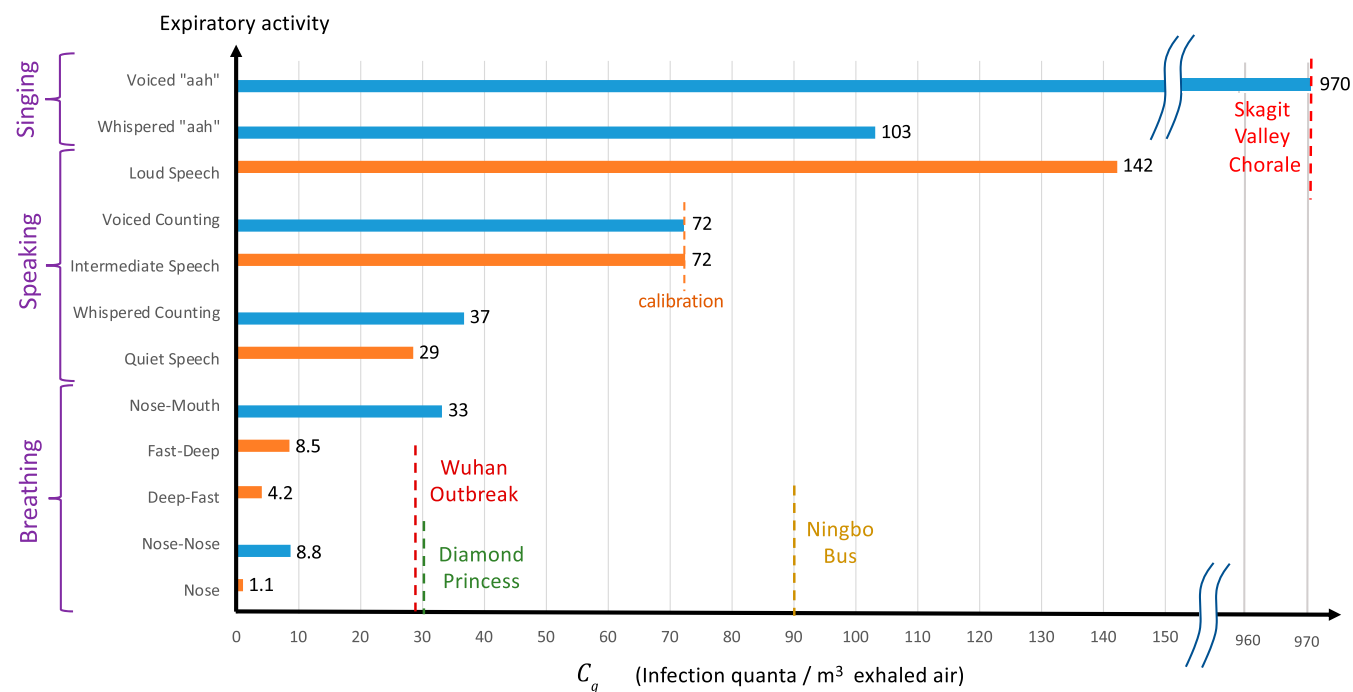


Fig. 2. Estimates of the "infectiousness" of exhaled air, C_q , defined as the peak concentration of COVID-19 infection quanta in the breath of an infected person, for various respiratory activities. Values are deduced from the drop size distributions reported by Morawska et al. (11) (blue bars) and Asadi et al. (39) (orange bars). The only value reported in the epidemiological literature, $C_q = 970$ quanta/m³, was estimated (25) for the Skagit Valley Chorale superspreading event (27), which we take as a baseline case ($s_r = 1$) of elderly individuals exposed to the original strain of SARS-CoV-2. This value is rescaled by the predicted infectious aerosol volume fractions, $\phi_1 = \int_0^{r_c} \phi_s(r) dr$, obtained by integrating the steady-state size distributions reported in Fig. 1 for different expiratory activities (11). Aerosol volume fractions calculated for various respiratory activities from figure 5 of Asadi et al. (39) are rescaled so that the value $C_q = 72$ quanta/m³ for "intermediate speaking" matches that inferred from Morawska et al.'s (11) for "voiced counting." Estimates of C_q for the outbreaks during the quarantine period of the *Diamond Princess* (26) and the Ningbo bus journey (28), as well as the initial outbreak in Wuhan City (2, 81), are also shown (see *SI Appendix* for details).

high during the first week of symptoms and reaches 7×10^8 RNA copies per throat swab (92) (typically 1 mL to 3 mL). Since viral loads are 20 to 50% greater in sputum than in throat swabs (93), the most infectious aerosols are likely to contain $c_v \approx 10^9$ RNA copies per milliliter. Using this viral load and assuming $c_i = 2\%$ based on previous inferences for SARS-CoV (94), Buonanno et al. (17) estimated $c_q = 2 \times 10^7$ quanta/mL for SARS-CoV-2, an order of magnitude below our inferences obtained directly from spreading data for COVID-19 (11, 39). The inference that SARS-CoV-2 is 10 times more infectious than SARS-CoV, with $c_i \approx 10\%$ (an infectious dose on the order of 10 aerosol-borne virions), is consistent with the fact that only the former caused a pandemic.

Our findings are consistent with emerging virological (3, 31, 66, 67) and epidemiological (5, 19, 23, 28, 29) evidence that SARS-CoV-2 is present and extremely infectious in respiratory aerosols and that indoor airborne transmission is the dominant driver of the COVID-19 pandemic (4, 22). Further support for this hypothesis is provided by crudely applying our indoor transmission model to a number of slightly less well characterized spreading events, as detailed in *SI Appendix*, all of which yield roughly consistent values of C_q (shown in Fig. 2). For the initial outbreak of COVID-19 in Wuhan City (2, 81), we assume that spreading occurred predominantly in family apartments, as is consistent with the inference that 80% of transmission clusters arose in people's homes (32). We may then tentatively equate the average reproduction number estimated for the Wuhan outbreak (81), $\mathcal{R}_0 = 3.3$, with the indoor reproduction number, $\mathcal{R}_{in}(\tau)$. We use $\tau = 5.5$ d as the exposure time, assuming that it corresponds to mean time before the onset of symptoms and patient isolation. We consider the mean household size of three persons in a typical apartment with area 30 m^2 per person and a winter bedroom ventilation rate of 0.34 ACH (55), and assume that $\lambda_v = 0.3/\text{h}$ and $\bar{r} = 2 \text{ }\mu\text{m}$. We thus infer $C_q = 30 \text{ quanta/m}^3$, a value expected for normal breathing (Fig. 2).

For the Ningbo bus incident, all model parameters are known except for the air exchange rate. We estimate $\lambda_a = 1.25/\text{h}$ for a moving bus with closed windows, based on studies of pollutants in British transit buses (99). We thus infer $C_q = 90 \text{ quanta/m}^3$, a value that lies in the range of intermediate speaking, as might be expected onboard a bus filled to capacity. Considering the uncertainty in λ_a , one might also infer a value consistent with resting on a quiet bus; in particular, choosing $\lambda_a = 0.34/\text{h}$ yields $C_q = 57 \text{ quanta/m}^3$. Finally, we infer a value of $C_q = 30 \text{ quanta/m}^3$ from the spreading event onboard the quarantined *Diamond Princess* cruise ship (26), a value consistent with the passengers being primarily at rest. However, we note that the extent to which the *Diamond Princess* can be adequately described in terms of a well-mixed space remains the subject of some debate (see *SI Appendix*, section 5).

We proceed by making the simplifying assumption that the dependence of C_q on expiratory activity illustrated in Fig. 2 is universal, but retain the freedom to rescale these values by the relative transmissibility s_r for different age groups and viral strains. It is well established that children have considerably lower hospitalization and death rates (86–88), but there is growing evidence that they also have lower transmissibility (89–91, 100, 101). A recent study of household clusters suggests that children are rarely index cases or involved in secondary transmissions (89). The best controlled comparison comes from quarantined households in China, where social contacts were reduced sevenfold to eightfold during lockdowns (101). Compared to the elderly (over 65 y old) for which we have assigned $s_r = 1$, the relative susceptibility of adults (aged 15 y to 64 y) was found to be $s_r = 68\%$, while that of children (aged 0 y to 14 y) was $s_r = 23\%$. We proceed by using these values of s_r for these

three different age groups and the original strain of SARS-CoV-2 in our case studies. However, we anticipate the need to revise these s_r values for new viral variants, such as the lineage B.1.1.7 (VOC 202012/01) (84, 85), which recently emerged in the United Kingdom with 60% greater transmissibility and elevated risk of infection among children.

In summary, our inferences of C_q and s_r from a diverse set of indoor spreading events and from independent physiological data are sufficiently self-consistent to indicate that the values reported in Fig. 2 may prove to be sufficient to apply the safety guideline in a quantitative fashion. Our hope is that our attempts to infer C_q will motivate the collection of more such data from spreading events, which might then be used to refine our necessarily crude initial estimates.

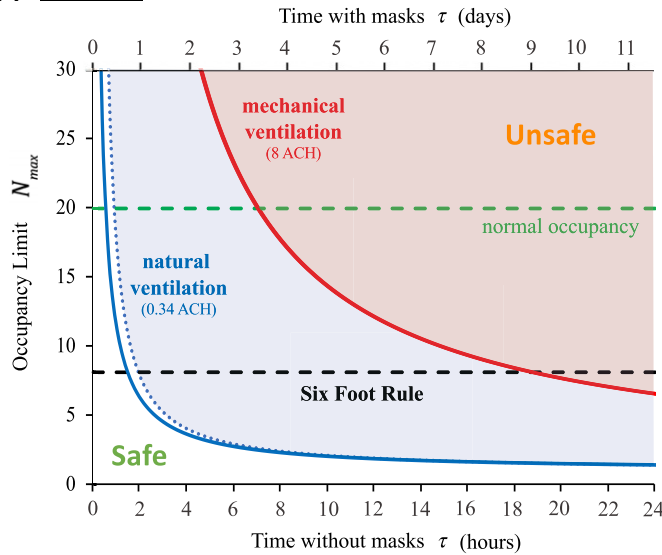
Case Studies

We proceed by illustrating the value of our guideline in estimating the maximum occupancy or exposure time in two settings of particular interest, the classroom and an elder care facility. Considering our inferences from the data and the existing literature, it would appear reasonable to illustrate our guideline for COVID-19 with the conservative choice of $C_q = 30 \text{ quanta/m}^3$. However, we emphasize that this value is expected to vary strongly with different demographics and respiratory activity levels (17). In taking the value of $C_q = 30 \text{ quanta/m}^3$, we are assuming that, in both settings considered, occupants are engaged in relatively mild respiratory activities consistent with quiet speech or rest. In assessing critical CETs for given populations, we stress that the tolerance ϵ is a parameter that should be chosen judiciously according to the vulnerability of the population, which varies dramatically with age and preexisting conditions (86–89).

We first apply our guideline to a typical American classroom, designed for an occupancy of 19 students and their teacher, and choose a modest risk tolerance, $\epsilon = 10\%$ (Fig. 3A). The importance of adequate ventilation and mask use is made clear by our guideline. For normal occupancy and without masks, the safe time after an infected individual enters the classroom is 1.2 h for natural ventilation and 7.2 h with mechanical ventilation, according to the transient bound, *SI Appendix*, Eq. S8. Even with cloth mask use ($p_m = 0.3$), these bounds are increased dramatically, to 8 and 80 h, respectively. Assuming 6 h of indoor time per day, a school group wearing masks with adequate ventilation would thus be safe for longer than the recovery time for COVID-19 (7 d to 14 d), and school transmissions would be rare. We stress, however, that our predictions are based on the assumption of a “quiet classroom” (38, 77), where resting respiration ($C_q = 30$) is the norm. Extended periods of physical activity, collective speech, or singing would lower the time limit by an order of magnitude (Fig. 2).

Our analysis sounds the alarm for elderly homes and long-term care facilities, which account for a large fraction of COVID-19 hospitalizations and deaths (86–88). In nursing homes in New York City, law requires a maximum occupancy of three and recommends a minimum area of 80 ft^2 per person. In Fig. 3B, we plot the guideline for a tolerance of $\epsilon = 0.01$ transmission probability, chosen to reflect the vulnerability of the community. Once again, the effect of ventilation is striking. For natural ventilation (0.34 ACH), the Six-Foot Rule fails after only 3 min under quasi-steady conditions, or after 17 min for the transient response to the arrival of an infected person, in which case the Fifteen-Minute Rule is only marginally safe. With mechanical ventilation (at 8 ACH) in steady state, three occupants could safely remain in the room for no more than 18 min. This example provides insight into the devastating toll of the COVID-19 pandemic on the elderly (86, 88). Furthermore, it underscores the need to minimize the sharing of indoor space, maintain

A Classroom



B Nursing home

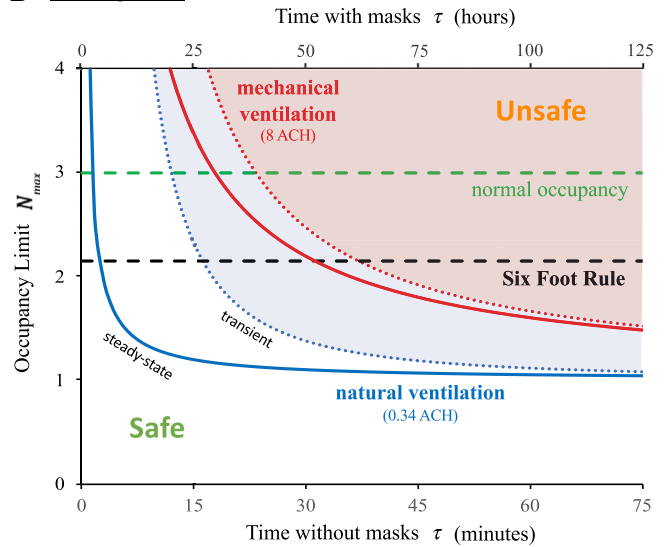


Fig. 3. The COVID-19 indoor safety guideline would limit the cumulative exposure time (CET) in a room with an infected individual to lie beneath the curves shown. Solid curves are deduced from the pseudo-steady formula, Eq. 5, for both natural ventilation ($\lambda_a = 0.34/h$; blue curve) and mechanical ventilation ($\lambda_a = 8.0/h$; red curve). Horizontal axes denote occupancy times with and without masks. Evidently, the Six-Foot Rule (which limits occupancy to $N_{max} = \sqrt{A}/(6 \text{ ft})$) becomes inadequate after a critical time, and the Fifteen-Minute Rule becomes inadequate above a critical occupancy. (A) A typical school classroom: 20 persons share a room with an area of 900 ft^2 and a ceiling height of 12 ft ($A = 83.6 \text{ m}^2$, $V = 301 \text{ m}^3$). We assume low relative transmissibility ($s_r = 25\%$), cloth masks ($p_m = 30\%$), and moderate risk tolerance ($\epsilon = 10\%$) suitable for children. (B) A nursing home shared room ($A = 22.3 \text{ m}^2$, $V = 53.5 \text{ m}^3$) with a maximum occupancy of three elderly persons ($s_r = 100\%$), disposable surgical or hybrid-fabric masks ($p_m = 10\%$), and a lower risk tolerance ($\epsilon = 1\%$) to reflect the vulnerability of the community. The transient formula, SI Appendix, Eq. S8, is shown with dotted curves. Other parameters are $C_q = 30 \text{ quanta/m}^3$, $\lambda_v = 0.3/h$, $Q_b = 0.5 \text{ m}^3/h$, and $\bar{r} = 0.5 \mu\text{m}$.

adequate, once-through ventilation, and encourage the use of face masks.

In both examples, the benefit of face masks is immediately apparent, since the CET limit is enhanced by a factor p_m^{-2} , the inverse square of the mask penetration factor. Standard surgical masks are characterized by $p_m = 1$ to 5% (73, 74), and so allow the CET to be extended by 400 to 10,000 times. Even cloth face coverings would extend the CET limit by 6 to 100 times for hybrid fabrics ($p_m = 10$ to 40%) or 1.5 to 6 times for single-layer fabrics ($p_m = 40$ to 80%) (75). Our inference of the efficacy of face masks in mitigating airborne transmission is roughly consistent with studies showing the benefits of mask use on COVID-19 transmission at the scales of both cities and countries (22, 33, 83).

Air filtration has a less dramatic effect than face mask use in increasing the CET bound. Nevertheless, it does offer a means of mitigating indoor transmission with greater comfort, albeit at greater cost (22, 72). Eq. 5 indicates that even perfect air filtration, $p_f = 1$, will only have a significant effect in the limit of highly recirculated air, $Z_p \ll 1$. The corresponding minimum outdoor airflow per person, Q/N_{max} , should be compared with local standards, such as 3.8 L/s per person for retail spaces and classrooms and 10 L/s per person for gyms and sports facilities (72). In the above classroom example with a typical primary outdoor air fraction of $Z_p = 20\%$ (22), the air change rate λ_a could effectively be increased by a factor of 4.6 by installing a MERV-13 filter, $p_m = 90\%$, or a factor of 5.0 with a HEPA filter, $p_m = 99.97\%$. At high air exchange rates, the same factors would multiply the CET bound.

Next, we illustrate the value of our guideline in contact tracing (82), specifically, in prescribing the scope of the testing of people with whom an infected index case has had close contact. The CDC presently defines a COVID-19 “close contact” as any encounter in which an individual is within 6 ft of an infected person for more than 15 min. Fig. 3 makes clear that this definition may grossly underestimate the number of individuals exposed

to a substantial risk of airborne infection in indoor spaces. Our study suggests that, whenever our CET bound (5) is violated during an indoor event with an infected person, at least one transmission is likely, with probability ϵ . When the tolerance ϵ exceeds a critical value, all occupants of the room should be considered close contacts and so warrant testing. For relatively short exposures ($\lambda_a \tau \ll 1$) initiated when the index case enters the room, the transient bound should be considered (SI Appendix, section 2).

We proceed by considering the implications of our guideline for the implementation of quarantining and testing. While official quarantine guidelines emphasize the importance of isolating infected persons, our study makes clear the importance of isolating and clearing infected indoor air. In cases of home quarantine of an infected individual with healthy family members, our guideline provides specific recommendations for mitigating indoor airborne transmission. For a group sharing an indoor space intermittently, for example, office coworkers or classmates, regular testing should be done with a frequency that ensures that the CET between tests is less than the limit set by the guideline. Such testing would become unnecessary if the time limit set by the CET bound greatly exceeds the time taken for an infected person to be removed from the population. For the case of a symptomatic infected person, this removal time should correspond to the time taken for the onset of symptoms ($\sim 5.5 \text{ d}$). To safeguard against asymptomatic individuals, one should use the recovery time ($\sim 14 \text{ d}$) in place of the removal time.

Finally, we briefly discuss how the prevalence of infection in the population affects our safety guideline. Our guideline sets a limit on the indoor reproductive number, the risk of transmission from a single infected person in the room. It thus implicitly assumes that the prevalence of infection in the population, p_i , is relatively low. In this low- p_i limit, the risk of transmission increases with the expected number of infected persons in the

room, Np_i , and the tolerance should be lowered in proportion to Np_i if it exceeds one. Conversely, when $Np_i \rightarrow 0$, the tolerance might be increased proportionally until the recommended restrictions are deemed unnecessary.

For instructions on how to apply our guideline to other situations, we refer the interested reader to the spreadsheet provided in *SI Appendix*. There, by specifying a given room geometry, ventilation rate, and respiratory activity, one may deduce the maximum CET in a particular indoor setting, and so define precisely what constitutes an exposure in that setting. An online app based on our guideline has also been developed (102).

Beyond the Well-Mixed Room

The model developed herein describes the risk of small respiratory drops ($r < r_c$) in the case where the entirety of the room is well mixed. There are undoubtedly circumstances where there are substantial spatial and temporal variations of the pathogen concentration from the mean (7, 42). For example, it is presumably the spatial variations from well mixedness that result in the inhomogeneous infection patterns reported for a number of well-documented transmission events in closed spaces, including a COVID outbreak in a Chinese restaurant (4), and SARS outbreaks on airliners (103). Circumstances have also been reported where air conditioner-induced flows appear to have enhanced direct pathogen transport between infected and susceptible individuals (104). In the vicinity of an infected person, the turbulent respiratory jet or puff will have a pathogen concentration that is substantially higher than the ambient (20, 43). Chen et al. (42) referred to infection via respiratory plumes as “short-range airborne transmission,” and demonstrated that it poses a substantially greater risk than large-drop transmission. In order to distinguish short-range airborne transmission from that considered in our study, we proceed by referring to the latter as “long-range airborne” transmission.

On the basis of the relatively simple geometric form of turbulent jet and puff flows, one may make estimates of the form of the mixing that respiratory outflows induce, the spatial distribution of their pathogen concentration, and so the resulting risk they pose to the room’s occupants. For the case of the turbulent jet associated with relatively continuous speaking or breathing, turbulent entrainment of the ambient air leads to the jet radius $r = \alpha_t x$ increasing linearly with distance x from the source, where $\alpha_t \approx 0.1$ to 0.15 is the typical jet entrainment coefficient (20, 42, 43). The conservation of momentum flux $M = \pi \rho_a r^2 v^2$ then indicates that the jet speed decreases with distance from the source according to $v(x) = M^{1/2} / (\alpha_t x \sqrt{\pi \rho_a})$. Concurrently, turbulent entrainment results in the pathogen concentration within the jet decreasing according to $C_j(x)/C_0 = A_m^{1/2} / (\alpha_t x)$, where $A_m \approx 2 \text{ cm}^2$ denotes the cross-sectional area of the mouth, and $C_0 = C_q/c_v$ is the exhaled pathogen concentration.[§] Abkarian et al. (43) thus deduce that, for the respiratory jet generated by typical speaking, the concentration of pathogen is diminished to approximately 3% of its initial value at a distance of 2 m.

In a well-mixed room, the mean concentration of pathogen produced by a single infected person is $f_d C_0$. For example, in the large, poorly ventilated room of the Skagit Valley Chorale, we compute a dilution factor, $f_d = Q_b / (\lambda_c(\bar{r}) V)$, of approximately 0.001. We note that, since $\lambda_c(r) > \lambda_a = Q/V$, the dilution factor satisfies the bound, $f_d \leq Q_b/Q$. For typical rooms and air exchange rates, f_d lies in the range of 0.0001 to 0.01. With

the dilution factor of the well-mixed room and the dilution rate of respiratory jets, we may now assess the relative risk to a susceptible person of a close encounter (either episodic or prolonged) with an infected individual’s respiratory jet, and an exposure associated with sharing a room with an infected person for an extended period. Since the infected jet concentration $C_j(x)$ decreases with distance from its source, one may assess its pathogen concentration relative to that of the well-mixed room, $C_j(x)/(f_d C_0) = A_m^{1/2} / (\alpha_t f_d x)$. There is thus a critical distance, $A_m^{1/2} / (\alpha_t f_d)$, beyond which the pathogen concentration in the jet is reduced to that of the ambient. This distance exceeds 10 m for f_d in the aforementioned range and so is typically much greater than the characteristic room dimension. Thus, in the absence of masks, respiratory jets may pose a substantially greater risk than the well-mixed ambient.

We first consider a worst-case, close-contact scenario in which a person directly ingests a lung full of air exhaled by an infected person. An equivalent amount of pathogen would be inhaled from the ambient by anyone within the room after a time $\tau = V_b / (Q_b f_d)$, where $V_b \approx 500 \text{ mL}$ is the volume per breath. For the geometry of the Skagit choir room, for which $f_d = 0.001$, the critical time beyond which airborne transmission is a greater risk than this worst-case close encounter with a respiratory plume is $\tau = 1.0 \text{ h}$. We next consider the worst-case scenario governed by the Six-Foot Rule, in which a susceptible person is directly in the path of an infected turbulent jet at a distance of 6 ft, over which the jet is diluted by a factor of 3% (43). The associated concentration in the jet is still roughly 30 times higher than the steady-state concentration in the well-mixed ambient (when $f_d = 0.001$), and so would result in a commensurate amplification of the transmission probability. Our guideline could thus be adopted to safeguard against the risk of respiratory jets in a socially distanced environment by reducing ϵ by a factor of $C(6\text{ft})/(f_d C_0)$, which is 3 to 300 for f_d in the range of 0.0001 to 0.01. We note that the latter worst-case scenario describes a static situation where a susceptible individual is seated directly in the respiratory plume of an infected individual, as may arise in a classroom or airplane (103). More generally, with a circulating population in an indoor setting, one would expect to encounter an infected respiratory plume only for some small fraction of the time, consideration of which would allow for a less conservative choice of ϵ .

We may thus make a relatively crude estimate for the additional risk of short-range plume transmission, appropriate when masks are not being worn ($p_m = 1$), by adding a correction to our safety guideline [5]. We denote by p_j the probability that a susceptible neighbor lies in the respiratory plume of the infected person, and denote by $x > 0$ the distance between nearest neighbors, between which the risk of infection is necessarily greatest. We thus deduce

$$\mathcal{R}_{in}(\tau) \left[1 + \frac{p_j A_m^{1/2}}{N_s f_d \alpha_t x} \right] < \epsilon. \quad [7]$$

In certain instances, meaningful estimates may be made for both p_j and x . For example, if a couple dines at a restaurant, x would correspond roughly to the distance across a table, and p_j would correspond to the fraction of the time they face each another. If N occupants are arranged randomly in an indoor space, then one expects $p_j = \tan^{-1} \alpha_t / \pi$ and $x = \sqrt{A/N}$. When strict social distancing is imposed, one may further set x to the minimum allowed interperson distance, such as 6 ft. Substitution from Eq. 5 reveals that the second term in Eq. 7 corresponds to the risk of transmission from respiratory jets, as deduced by Yang et al. (106), aside from the factor p_j . We note that any such guideline intended to mitigate against short-range airborne transmission by respiratory plumes will be, as is [7], dependent on geometry,

[§]These expressions for $v(x)$ and $C(x)$ are valid in the limit of $x > x_v$, where x_v is the virtual origin of the jet, typically on the order of 10 cm (20, 105). Near-field expressions well behaved at $x = 0$ are given by replacing x with $x + x_v$, and normalizing such that $C(0) = C_0$.

flow, and human behavior, while our guideline for the mitigation of long-range airborne transmission [5] is universal.

We note that the use of face masks will have a marked effect on respiratory jets, with the fluxes of both exhaled pathogen and momentum being reduced substantially at their source. Indeed, Chen et al. (42) note that, when masks are worn, the primary respiratory flow may be described in terms of a rising thermal plume, which is of significantly less risk to neighbors. With a population of individuals wearing face masks, the risk posed by respiratory jets will thus be largely eliminated, while that of the well-mixed ambient will remain.

Finally, we stress that our guideline is based on the average concentration of aerosols within the room. For every region of enhanced airborne pathogen concentration, there is necessarily a region of reduced concentration and lower transmission risk elsewhere in the room. The ensemble average of the transmission risk over a number of similar events, and the time-averaged transmission risk in a single event, are both expected to approach that in the well-mixed steady state, as in ergodic processes in statistical mechanics. This feature of the system provides rationale for the self-consistency of our inferences of C_q , based on the hypothesis of the well-mixed room, from the diverse set of spreading events considered herein.

Discussion and Caveats

We have focused here primarily on airborne transmission, for which infection arises through inhalation of a critical quantity of airborne pathogen, and neglected the roles of both contact and large-drop transmission (6). While motivated by the COVID-19 pandemic, our theoretical framework applies quite generally to airborne respiratory illnesses, including influenza. Moreover, we note that the approach taken, coupling the droplet dynamics to the transmission dynamics, allows for a more complete description. For example, consideration of conservation of pathogen allows one to calculate the rate of pathogen sedimentation and associated surface contamination, consideration of which would allow for quantitative models of contact transmission and so inform cleaning protocols.

Typical values for the parameters arising in our model are listed in *SI Appendix, Table S1*. Respiration rates Q_b have been measured to be $\sim 0.5 \text{ m}^3/\text{h}$ for normal breathing, and may increase by a factor of 3 for more strenuous activities (17). Other parameters, including room geometry, ventilation, and filtration rates, will obviously be room dependent. The most poorly constrained parameter appearing in our guideline is $C_q s_r$, the product of the concentration of pathogen in the breath of an infected person and the relative transmissibility. The latter, s_r , was introduced in order to account for the dependence of transmissibility on the mean age of the population (86–88, 91) and the viral strain (84, 85). The value of $C_q s_r$ was inferred from the best characterized superspreading event, the Skagit Valley Chorale incident (25), as arose among an elderly population with a median age of 69 y (27), for which we assign $s_r = 1$. The C_q value so inferred was rescaled using reported drop size distributions (11, 23, 38) allowing us to estimate C_q for several respiratory activities, as listed in Fig. 3. Further comparison with inferences based on other spreading events of new viral strains among different populations would allow for refinement of our estimates of C_q and s_r . We thus appeal to the public health community to document the physical conditions enumerated in *SI Appendix, Table S1* for more indoor spreading events.

Adherence to the Six-Foot Rule would limit large-drop transmission, and adherence to our guideline, Eq. 5, would limit long-range airborne transmission. We have also shown how the sizable variations in pathogen concentration associated with respiratory flows, arising in a population not wearing face masks, might be taken into account. Consideration of both short-range

and long-range airborne transmission leads to a guideline of the form of Eq. 7 that would bound both the distance between occupants and the CET. Circumstances may also arise where a room is only partially mixed, owing to the absence or deficiency of air conditioning and ventilation flows, or the influence of irregularities in the room geometry (107). For example, in a poorly ventilated space, contaminated warm air may develop beneath the ceiling, leading to the slow descent of a front between relatively clean and contaminated air, a process described by “filling-box” models (107). In the context of reducing COVID-19 transmission in indoor spaces, such variations from well-mixedness need be assessed on a room-by-room basis. Nevertheless, the criterion [5] represents a minimal requirement for safety from long-range airborne infection in well-mixed, indoor spaces.

We emphasize that our guideline was developed specifically with a view to mitigating the risk of long-range airborne transmission. We note, however, that our inferences of C_q came from a number of superspreading events, where other modes of transmission, such as respiratory jets, are also likely to have contributed. Thus, our estimates for C_q are necessarily overestimates, expected to be higher than those that would have arisen from purely long-range airborne transmission. Consequently, our safety guideline for airborne transmission necessarily provides a conservative upper bound on CET. We note that the additional bounds required to mitigate other transmission modes will not be universal; for example, we see, in Eq. 7, that the danger of respiratory jets will depend explicitly on the arrangement of the room’s occupants. Finally, we reiterate that the wearing of masks largely eliminates the risk of respiratory jets, and so makes the well-mixed room approximation considered here all the more relevant.

Our theoretical model of the well-mixed room was developed specifically to describe airborne transmission between a fixed number of individuals in a single well-mixed room. Nevertheless, we note that it is likely to inform a broader class of transmission events. For example, there are situations where forced ventilation mixes air between rooms, in which case the compound room becomes, effectively, a well-mixed space. Examples considered here are the outbreaks on the *Diamond Princess* and in apartments in Wuhan City (see *SI Appendix*); others would include prisons. There are many other settings, including classrooms and factories, where people come and go, interacting intermittently with the space, with infected people exhaling into it, and susceptible people inhaling from it, for limited periods. Such settings are also informed by our model, provided one considers the mean population dynamics, and so identifies N with the mean number of occupants.

The guideline [5] depends on the tolerance ϵ , whose value in a particular setting should be set by the appropriate policy makers, informed by the latest epidemiological evidence. Likewise, the guideline includes the relative transmissibility s_r of a given viral strain within a particular subpopulation. These two factors may be eliminated from consideration by using [6] to assess the relative behavioral risk posed to a particular individual by attending a specific event of duration τ with N other participants. We thus define a relative risk index,

$$I_R = \frac{N\tau C_q Q_b^2 p_m^2}{\lambda_a V}, \quad [8]$$

that may be evaluated using appropriate C_q and Q_b values (listed in *SI Appendix, Table S2*). One’s risk increases linearly with the number of people in a room and duration of the event. Relative risk decreases for large, well-ventilated rooms and increases when the room’s occupants are exerting themselves or speaking loudly. While these results are intuitive, the approach taken here provides a physical framework for understanding them

quantitatively. It also provides a quantitative measure of the relative risk of certain environments, for example, a well-ventilated, sparsely occupied laboratory and a poorly ventilated, crowded, noisy bar. Along similar lines, the weighted average of [8], provides a quantitative assessment of one's risk of airborne infection over an extended period. It thus allows for a quantitative assessment of what constitutes an exposure, a valuable notion in defining the scope of contact tracing, testing, and quarantining.

Above all, our study makes clear the inadequacy of the Six-Foot Rule in mitigating indoor airborne disease transmission, and offers a rational, physically informed alternative for managing life in the time of COVID-19. If implemented, our safety guideline would impose a limit on the CET in indoor settings, violation of which constitutes an exposure for all of the room's occupants. Finally, while our study has allowed for an estimate of the infectiousness of COVID-19, it also indicates how new data characterizing indoor spreading events may lead to improved

estimates thereof and so to quantitative refinements of our safety guideline.

The spreadsheet included in [Dataset S1](#) provides a simple means of evaluating the CET limit for any particular indoor setting. A convenient online app based on our safety guideline is also available (102). The app and spreadsheet also enable the use of data from CO₂ sensors (47) to improve the accuracy of the safety guideline (108). A glossary of terms arising in our study is presented in [SI Appendix, Table S3](#).

Data Availability. All study data are included in the article and supporting information.

ACKNOWLEDGMENTS. We thank William Ristenpart and Sima Asadi for sharing experimental data, and Lesley Bazant, Lydia Bourouiba, Daniel Cogswell, Mark Hampden-Smith, Kyle Hofmann, David Keating, Lidia Morawska, Nels Olson, Monona Rossol, and Renyi Zhang for important references.

1. N. Chen *et al.*, Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: A descriptive study. *Lancet* **395**, 507–513 (2020).
2. Q. Li *et al.*, Early transmission dynamics in Wuhan, China, of novel coronavirus-infected pneumonia. *N. Engl. J. Med.* **382**, 1199–1207 (2020).
3. J. A. Lednicky *et al.*, Viable SARS-CoV-2 in the air of a hospital room with COVID-19 patients. *Int. J. Infect. Dis.* **100**, 476–482 (2020).
4. L. Morawska, D. K. Milton, It is time to address airborne transmission of COVID-19. *Clin. Infect. Dis.* **71**, 2311–2313 (2020).
5. L. Morawska, J. Cao, Airborne transmission of SARS-CoV-2: The world should face the reality. *Environ. Int.* **139**, 105730 (2020).
6. R. Mittal, R. Ni, J.-H. Seo, The flow physics of COVID-19. *J. Fluid Mech.* **894**, F2 (2020).
7. M. Jayaweera, H. Perera, B. Gunawardana, J. Manatunge, Transmission of COVID-19 virus by droplets and aerosols. *Environ. Res.* **188**, 109819 (2020).
8. L. Morawska, Droplet fate in indoor environments, or can we prevent the spread of infection? *Indoor Air* **16**, 335–347 (2006).
9. P. Khare, L. C. Marr, Simulation of vertical concentration gradient of influenza viruses in dust resuspended by walking. *Indoor Air* **25**, 428–440 (2015).
10. S. Asadi *et al.*, Influenza A virus is transmissible via aerosolized fomites. *Nat. Commun.* **11**, 4062 (2020).
11. L. Morawska *et al.*, Size distribution and sites of origin of droplets expelled from the human respiratory tract during expiratory activities. *J. Aerosol Sci.* **40**, 256–269 (2009).
12. L. Gammaitoni, M. C. Nucci, Using a mathematical model to evaluate the efficacy of TB control measures. *Emerg. Infect. Dis.* **3**, 335 (1997).
13. C. B. Beggs, C. J. Noakes, P. A. Sleight, L. A. Fletcher, K. Siddiqi, The transmission of tuberculosis in confined spaces: An analytical review of alternative epidemiological models. *Int. J. Tubercul. Lung Dis.* **7**, 1015–1026 (2003).
14. M. Nicas, W. W. Nazaroff, A. Hubbard, Toward understanding the risk of secondary airborne infection: Emission of respirable pathogens. *J. Occ. Env. Hygiene* **2**, 143–154 (2005).
15. C. J. Noakes, C. B. Beggs, P. A. Sleight, K. G. Kerr, Modelling the transmission of airborne infections in enclosed spaces. *Epidemiol. Infect.* **134**, 1082–1091 (2006).
16. N. I. Stilianakis, Y. Drossinos, Dynamics of infectious disease transmission by inhalable respiratory droplets. *J. R. Soc. Interface* **7**, 1355–1366 (2010).
17. G. Buonanno, L. Stabile, L. Morawska, Estimation of airborne viral emission: Quanta emission rate of SARS-CoV-2 for infection risk assessment. *Environ. Int.* **141**, 105794 (2020).
18. G. Buonanno, L. Morawska, L. Stabile, Quantitative assessment of the risk of airborne transmission of SARS-CoV-2 infection. *Environ. Int.* **145**, 106112 (2020).
19. L. Setti *et al.*, Airborne transmission route of COVID-19: Why 2 meters/6 ft of interpersonal distance could not be enough. *Int. J. Environ. Res. Publ. Health* **17**, 2932 (2020).
20. L. Bourouiba, E. Dehandschoewercker, J. W. M. Bush, Violent expiratory events: On coughing and sneezing. *J. Fluid Mech.* **745**, 537–563 (2014).
21. R. S. Papineni, F. S. Rosenthal, The size distribution of droplets in the exhaled breath of healthy human subjects. *J. Aerosol Med.* **10**, 105–116 (1997).
22. R. Zhang, Y. Li, A. L. Zhang, Y. Wang, M. J. Molina, Identifying airborne transmission as the dominant route for the spread of COVID-19. *Proc. Natl. Acad. Sci. U.S.A.* **117**, 14857–14863 (2020).
23. S. Asadi, N. Bouvier, A. S. Wexler, W. D. Ristenpart, The coronavirus pandemic and aerosols: Does COVID-19 transmit via expiratory particles? *Aerosol. Sci. Technol.* **54**, 635–638 (2020).
24. K. A. Prather *et al.*, Airborne transmission of SARS-CoV-2. *Science* **370**, 303–304 (2020).
25. S. L. Miller *et al.*, Transmission of SARS-CoV-2 by inhalation of respiratory aerosol in the Skagit Valley Chorale superspreading event. *Indoor Air* **31**, 314–323 (2020).
26. L. F. Moriarty, Public health responses to COVID-19 outbreaks on cruise ships worldwide, February–March 2020. *MMWR. Morb. Mortal. Wkly. Rep.* **69**, 347–352 (2020).
27. L. Hamner, High SARS-CoV-2 attack rate following exposure at a choir practice, Skagit County, Washington, March 2020. *MMWR. Morb. Mortal. Wkly. Rep.* **69**, 606–610 (2020).
28. Y. Shen *et al.*, Community outbreak investigation of SARS-CoV-2 transmission among bus riders in eastern China. *JAMA Int. Med.* **180**, 1665–1671 (2020).
29. H. Nishiura *et al.*, Closed environments facilitate secondary transmission of coronavirus disease 2019 (COVID-19). medRxiv [Preprint] (2020). <https://doi.org/10.1101/2020.02.28.20029272> (Accessed 1 July 2020).
30. S. E. Hwang, J. H. Chang, O. Bumjo, J. Heo, Possible aerosol transmission of COVID-19 associated with an outbreak in an apartment in Seoul, South Korea. *Int. J. Infect. Dis.* **104**, 73–76 (2020).
31. J. L. Santarpia *et al.*, The infectious nature of patient-generated SARS-CoV-2 aerosol. medRxiv [Preprint] (2020). <https://doi.org/10.1101/2020.07.13.20041632> (Accessed 1 August 2020).
32. H. Qian *et al.*, Indoor transmission of SARS-CoV-2. *Indoor Air*, 10.1111/ina.12766 (2020).
33. R. O. J. H. Stutt, R. Retkute, M. Bradley, C. A. Gilligan, J. Colvin, A modelling framework to assess the likely effectiveness of facemasks in combination with lock-down in managing the COVID-19 pandemic. *Proc. R. Soc. A.* **476**, 20200376 (2020).
34. J. B. Grotberg, Respiratory fluid mechanics. *Phys. Fluids* **23**, 021301 (2011).
35. D. R. Johnson, L. Morawska, The mechanism of breath aerosol formation. *J. Aerosol Med. Pulm. Drug Deliv.* **22**, 229–237 (2009).
36. G. Johnson *et al.*, Modality of human expired aerosol size distributions. *J. Aerosol Sci.* **42**, 839–851 (2011).
37. M. Abkarian, H. A. Stone, Stretching and break-up of saliva filaments during speech: A route for pathogen aerosolization and its potential mitigation. *Phys. Rev. Fluids* **5**, 102301 (2020).
38. S. Asadi *et al.*, Aerosol emission and superemission during human speech increase with voice loudness. *Sci. Rep.* **9**, 2348 (2019).
39. S. Asadi *et al.*, Effect of voicing and articulation manner on aerosol particle emission during human speech. *PLoS One* **15**, e0227699 (2020).
40. B. E. Scharfman, A. H. Techet, J. W. M. Bush, L. Bourouiba, Visualization of sneeze ejecta: Steps of fluid fragmentation leading to respiratory droplets. *Exp. Fluid* **57**, 24 (2016).
41. J. Gralton, E. Tovey, M.-L. McLaws, W. D. Rawlinson, The role of particle size in aerosolised pathogen transmission: A review. *J. Infect.* **62**, 1–13 (2011).
42. W. Chen, N. Zhang, J. Wei, H.-L. Yen, Y. Li, Short-range airborne route dominates exposure of respiratory infection during close contact. *Build. Environ.* **176**, 106859 (2020).
43. M. Abkarian, S. Mendez, N. Xue, Y. Fan, H. A. Stone, Speech can produce jet-like transport relevant to asymptomatic spreading of virus. *Proc. Natl. Acad. Sci. U.S.A.* **117**, 25237–25245 (2020).
44. W. F. Wells, *Airborne Contagion and Air Hygiene: An Ecological Study of Droplet Infections* (Harvard University Press, 1955).
45. E. C. Riley, G. Murphy, R. L. Riley, Airborne spread of measles in a suburban elementary school. *Am. J. Epidemiol.* **107**, 421–432 (1978).
46. C.-M. Liao, C.-F. Chang, H.-M. Liang, A probabilistic transmission dynamic model to assess indoor airborne infection risks. *Risk Anal.* **25**, 1097–1107 (2005).
47. S. N. Rudnick, D. K. Milton, Risk of indoor airborne infection transmission estimated from carbon dioxide concentration. *Indoor Air* **13**, 237–245 (2003).
48. S. Zhu *et al.*, Ventilation and laboratory confirmed acute respiratory infection (ARI) rates in college residence halls in College Park, Maryland. *Environ. Int.* **137**, 105537 (2020).
49. M. E. Davis, R. J. Davis, *Fundamentals of Chemical Reaction Engineering* (Courier Corporation, 2012).
50. W. M. Deen, *Analysis of Transport Phenomena* (Oxford University Press, ed. 2, 2011).
51. J. Corner, E. D. Pendlebury, The coagulation and deposition of a stirred aerosol. *Proc. Phys. Soc. B* **64**, 645, 1951.
52. A. C. K. Lai, W. W. Nazaroff, Modeling indoor particle deposition from turbulent flow onto smooth surfaces. *J. Aerosol Sci.* **31**, 463–476 (2000).
53. K.-C. Cheng *et al.*, Modeling exposure close to air pollution sources in naturally ventilated residences: Association of turbulent diffusion coefficient with air change rate. *Environ. Sci. Technol.* **45**, 4016–4022 (2011).

54. D. Martin, R. Nokes, Crystal settling in a vigorously converting magma chamber. *Nature* **332**, 534–536 (1988).
55. J. Hou *et al.*, Air change rates in urban Chinese bedrooms. *Indoor Air* **29**, 828–839 (2019).
56. W. F. Wells *et al.*, On air-borne infection. Study II. droplets and droplet nuclei. *Am. J. Hyg.* **20**, 611–618 (1934).
57. X. Xie, Y. Li, A. T. Y. Chwang, P. L. Ho, W. H. Seto, How far droplets can move in indoor environments—Revisiting the wells evaporation–falling curve. *Indoor Air* **17**, 211–225 (2007).
58. R. R. Netz, Mechanisms of airborne infection via evaporating and sedimenting droplets produced by speaking. *J. Phys. Chem. B* **124**, 7093–7101 (2020).
59. J. V. Fahy, B. F. Dickey, Airway mucus function and dysfunction. *N. Engl. J. Med.* **363**, 2233–2247 (2010).
60. D. K. Milton *et al.*, Influenza virus aerosols in human exhaled breath: Particle size, culturability, and effect of surgical masks. *PLoS Pathog.* **9**, e1003205 (2013).
61. R. Wölfel *et al.*, Virological assessment of hospitalized patients with COVID-2019. *Nature* **581**, 465–469 (2020).
62. L. C. Marr, J. W. Tang, J. Van Mullekom, S. S. Lakdawala, Mechanistic insights into the effect of humidity on airborne influenza virus survival, transmission and incidence. *J. R. Soc. Interface* **16**, 20180298 (2019).
63. G. J. Harper, Airborne micro-organisms: Survival tests with four viruses. *Epidemiol. Infect.* **59**, 479–486 (1961).
64. W. Yang, L. C. Marr, Dynamics of airborne influenza viruses indoors and dependence on humidity. *PLoS One* **6**, e21481 (2011).
65. K. Lin, L. C. Marr, Humidity-dependent decay of viruses, but not bacteria, in aerosols and droplets follows disinfection kinetics. *Environ. Sci. Technol.* **54**, 1024–1032 (2019).
66. A. C. Fears *et al.*, Comparative dynamic aerosol efficiencies of three emergent coronaviruses and the unusual persistence of SARS-CoV-2 in aerosol suspensions. medRxiv [Preprint] (2020). <https://doi.org/10.1101/2020.04.13.20063784> (Accessed 15 July 2020).
67. N. Van Doremalen *et al.*, Aerosol and surface stability of SARS-CoV-2 as compared with SARS-CoV-1. *N. Engl. J. Med.* **382**, 1564–1567 (2020).
68. O. V. Pyankov, S. A. Bodnev, O. G. Pyankova, I. E. Agranovski, Survival of aerosolized coronavirus in the ambient air. *J. Aerosol Sci.* **115**, 158–163 (2018).
69. F. J. García de Abajo *et al.*, Back to normal: An old physics route to reduce SARS-CoV-2 transmission in indoor spaces. *ACS Nano* **14**, 7704–7713 (2020).
70. A. Schwartz *et al.*, Decontamination and reuse of N95 respirators with hydrogen peroxide vapor to address worldwide personal protective equipment shortages during the SARS-CoV-2 (COVID-19) pandemic. *Applied Biosafety* **25**, 67–70 (2020).
71. E. S. Mousavi, K. J. G. Pollitt, J. Sherman, R. A. Martinello, Performance analysis of portable HEPA filters and temporary plastic anterooms on the spread of surrogate coronavirus. *Bu. Environ.* **183**, 107186 (2020).
72. B. Blocken *et al.*, Can indoor sports centers be allowed to re-open during the COVID-19 pandemic based on a certificate of equivalence? *Bu. Environ.* **180**, 107022 (2020).
73. C.-C. Chen, K. Willeke, Aerosol penetration through surgical masks. *Am. J. Infect. Control.* **20**, 177–184 (1992).
74. T. Oberg, L. M. Brosseau, Surgical mask filter and fit performance. *Am. J. Infect. Control.* **36**, 276–282 (2008).
75. A. Konda *et al.*, Response to letters to the editor on aerosol filtration efficiency of common fabrics used in respiratory cloth masks: Revised and expanded results. *ACS Nano* **14**, 10764–10770 (2020).
76. Y. Li *et al.*, Transmission of communicable respiratory infections and facemasks. *J. Multidiscip. Healthc.* **1**, 17 (2008).
77. S. Asadi *et al.*, Efficacy of masks and face coverings in controlling outward aerosol particle emission from expiratory activities. *Sci. Rep.* **10**, 1–13 (2020).
78. J. Pan, C. Harb, W. Leng, L. C. Marr, Inward and outward effectiveness of cloth masks, a surgical mask, and a face shield. *Aerosol Sci. Technol.*, 10.1080/02786826.2021.1890687 (2021).
79. L. Nicolaou, T. A. Zaki, Characterization of aerosol Stokes number in 90° bends and idealized extrathoracic airways. *J. Aerosol Sci.* **102**, 105–127 (2016).
80. P. van den Driessche, Reproduction numbers of infectious disease models. *Infect. Dis. Model.* **2**, 288–303 (2017).
81. Y. Liu, A. A. Gayle, A. Wilder-Smith, J. Rocklöv, The reproductive number of COVID-19 is higher compared to SARS coronavirus. *J. Trav. Med.* **27**, 1–4 (2020).
82. L. Ferretti *et al.*, Quantifying SARS-CoV-2 transmission suggests epidemic control with digital contact tracing. *Science* **368**, eabb6936 (2020).
83. Y. Li, R. Zhang, J. Zhao, M. J. Molina, Understanding transmission and intervention for the COVID-19 pandemic in the United States. *Sci. Total Environ.* **748**, 141560 (2020).
84. N. G. Davies *et al.*, Estimated transmissibility and severity of novel SARS-CoV-2 variant of concern 202012/01 in England. *Science*, 10.1126/science.abb3055 (2021).
85. Erik. Volz *et al.*, Transmission of SARS-CoV-2 lineage B.1.1.7 in England: Insights from linking epidemiological and genetic data. medRxiv [Preprint] (2021). <https://doi.org/10.1101/2020.12.30.20249034> (Accessed 5 January 2021).
86. S. Richardson *et al.*, Presenting characteristics, comorbidities, and outcomes among 5700 patients hospitalized with COVID-19 in the New York City area. *J. Am. Med. Assoc.* **323**, 2052–2059 (05 2020).
87. N. G. Davies *et al.*, Age-dependent effects in the transmission and control of COVID-19 epidemics. *Nat. Med.* **26**, 1205–1211 (2020).
88. S. Garg, Hospitalization rates and characteristics of patients hospitalized with laboratory-confirmed coronavirus disease 2019: COVID-NET, 14 states, March 1–30, 2020. *MMWR. Morb. Mortal. Wkly. Rep.* **69**, 458–464 (2020).
89. Y. Zhu *et al.*, A meta-analysis on the role of children in SARS-CoV-2 in household transmission clusters. *Clin. Infect. Dis.*, 10.1093/cid/cia1825 (2020).
90. A. P. S. Munro, S. N. Faust, Children are not COVID-19 super spreaders: Time to go back to school. *Arch. Dis. Child.* **105**, 618–619 (2020).
91. M. Riediker, L. Morawska, Low exhaled breath droplet formation may explain why children are poor SARS-CoV-2 transmitters. *Aerosol. Air Q. Res.* **20**, 1513–1515 (2020).
92. R. Wölfel *et al.*, Virological assessment of hospitalized patients with COVID-2019. *Nature* **581**, 465–469 (2020).
93. Y. Pan, D. Zhang, P. Yang, L. L. M. Poon, Q. Wang, Viral load of SARS-CoV-2 in clinical samples. *Lancet Infect. Dis.* **20**, 411–412 (2020).
94. T. Watanabe, T. A. Bartrand, M. H. Weir, T. Omura, C. N. Haas, Development of a dose-response model for SARS coronavirus. *Risk Anal.* **30**, 1129–1138 (2010).
95. K. K.-W. To *et al.*, Temporal profiles of viral load in posterior oropharyngeal saliva samples and serum antibody responses during infection by SARS-CoV-2: An observational cohort study. *Lancet Infect. Dis.* **20**, 565–574 (2020).
96. E. Villermaux, Fragmentation versus cohesion. *J. Fluid Mech.* **898**, P1 (2020).
97. L. Zheng, J. Xu, F. Wu, W. Xu, Z. Long, Influences of ventilation modes on the coughing droplet dispersion process in a cruise cabin. *Chinese J. Ship Res.* **11**, 2 (2016).
98. S. Zheng *et al.*, Viral load dynamics and disease severity in patients infected with SARS-CoV-2 in Zhejiang province, China, January–March 2020: Retrospective cohort study. *Br. Med. J.* **369**, m1443 (2020).
99. W. W. Song, M. R. Ashmore, A. C. Terry, The influence of passenger activities on exposure to particles inside buses. *Atmos. Environ.* **43**, 6271–6278 (2009).
100. O. Miron, K.-H. Yu, R. Wilf-Miron, I. Kohane, N. Davidovitch, COVID-19 infections following physical school reopening. *Arch. Dis. Child.*, 10.1136/archdischild-2020-321018 (2020).
101. J. Zhang *et al.*, Changes in contact patterns shape the dynamics of the COVID-19 outbreak in China. *Science* **368**, 1481–1486 (2020).
102. K. Khan, J. W. M. Bush, M. Z. Bazant, COVID-19 indoor safety guideline. <https://indoor-covid-safety.herokuapp.com>. Accessed 6 April 2021.
103. S. J. Olsen *et al.*, Transmission of the severe acute respiratory syndrome on aircraft. *N. Engl. J. Med.* **349**, 2146–2422 (2003).
104. K.-S. Kwon *et al.*, Evidence of long-distance droplet transmission of SARS-CoV-2 by direct air flow in a restaurant in Korea. *J. Kor. Med. Sci.* **35**, e415 (2020).
105. F. Ciriello, G. R. Hunt, Analytical solutions and virtual origin corrections for forced, pure and lazy turbulent plumes based on a universal entrainment function. *J. Fluid Mech.* **893**, A12 (2020).
106. F. Yang, A. A. Pahlavan, S. Mendez, M. Abkarian, H. A. Stone, Towards improved social distancing guidelines: Space and time dependence of virus transmission from speech-driven aerosol transport between two individuals. *Phys. Rev. Fluids* **5**, 122501(R) (2020).
107. R. K. Bhagat, M. S. D. Wykes, S. B. Dalziel, P. F. Linden, Effects of ventilation on the indoor spread of COVID-19. *J. Fluid Mech.* **903**, F1 (2020).
108. M. Z. Bazant, *et al.*, Monitoring carbon dioxide to quantify the risk of indoor airborne transmission of COVID-19. medRxiv [Preprint] (2021). <https://doi.org/10.1101/2021.04.04.21254903> (Accessed 9 April 2021).